**Control**

Formaldehyde and ethanol

**Kaiserling Solution**

All amounts come from the article I have. Others I have read say to have 10-20 times the volume of the specimens used.

Kaiserling I Solution: 2 liter formalin, 425 g potassium acetate, 225 g potassium nitrate, DI up to 20 liters

Kaiserling II solution: Alcohol 95%

Rejuvenator solution: Pyridine 100 ml, Sodium hydrosulphite 100 gm, DI 4 liters (The article I have says this will help restore color that was lost from fixing with formalin but doesn’t say how long to leave specimens in this solution. Searching online says that the alcohol will help keep the color.)

Kaiserling III solution: Potassium acetate 300 g, Glycerine 6 liters, DI make up to 10 liters, thymol crystals

**BHT solution**

Empty gallon jugs/jars, plastic funnel, formalin, BHT (Butyl hydroxytoluene, Shell Chemical Co. Ionol-40), ice 🡪 keep cold, alcohol (to store the specimen)

10 % formalin solution, BHT 1:200 10% formalin 🡪 unsure how many liters this will make

**Glycerol/GAF**

If GAF used need: 80 ml acetone, 400 ml glycerol, 120 ml formaldehyde, 3400 ml FSW (total of 4 liters). The original method only placed the parasites in solution for 3 minutes. Also, the worms were placed on slides, so they were not placed in alcohol. We could try this method, but it would have a lot of trial and error. The authors also caution that this solution will cause colors to fade if left in it for too long.

The glycerol method: glycerol (unsure if anything else is needed since this method was used on already preserved specimens. We could fix specimens normally, then flush them with glycerol…but that seems like a waste of materials).

425 g potassium acetate (500 g - $128.95 – Fisher Scientific)

300 g Potassium acetate (500 g - $128.95 – Fisher Scientific)

225 g potassium nitrate (100 g - $90.65 –Fisher Scientific)

100ml Pyridine (1 litre - $296.13 – Fisher Scientific)

100 g Sodium hydrosulphite (500 g - $84.80 – Fisher Scientific)

?? Butyl hydroxytoluene (100 g - $37.96 – Fisher Scientific)